

## Isolation of Fungal Contaminants from Various Working Surface Areas of a Tertiary care Teaching Hospital, Amalapuram

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### Abstract

**INTRODUCTION:** Hospital environments, particularly those in tertiary care teaching hospitals, play a critical role in patient care and medical education. However, these environments can also serve as reservoirs for various microbial contaminants, including fungi, which pose significant risks to immunocompromised patients, healthcare workers, and visitors. This study aims to assess the incidence of fungal contaminants on various working surface areas in a tertiary care teaching hospital in Amalapuram. By identifying the types and prevalence of fungi present, it is possible to understand the environmental factors contributing to fungal contamination and implement necessary infection control measures.

**OBJECTIVES:** The principal goal of this study is to identify the presence of fungi on various working surface areas of a Tertiary care Teaching Hospital, Amalapuram.

**MATERIALS AND METHODS:** Moistened swabs were collected from various surface areas including bed rails, door handles, medical equipment, light switches, and nursing stations, ceilings, AC airvents, walls, IV stands etc... and were inoculated on SDA and the fungi were identified based on morphology.

**RESULTS:** A total of 100 swabs were collected from different areas of the hospital- different wards, OTs, all ICUs. Out of 100 swabs, 64 swabs showed growth on SDA slants. Out of 64 isolates, 23 isolates were *Aspergillus niger* (40%), 10 were *A.fumigatus* (15%), 20 were *Fusarium* (31%), 4 were *Curvularia* (6%), 2 were *Cladosporium* (3%), *Allescheria boydii* - 1(1%), *Exophiala jaenselmei*-1(1%), *Gliocladium* -1(1%), *Phoma* -1(1%), *Chaetomium* -1 (%).

**CONCLUSION:** Most of the patients in the hospitals are immune compromised. These fungal isolates in the hospital environment may become an additional economic burden to their present situation as most of them are multi drug resistant. So, measures to control hospital infection in the hospital environment mainly ICUs is very much needed

### INTRODUCTION:

Hospital environments, particularly those in tertiary teaching hospitals, play a critical role in patient care and medical education. However, these environments can also serve as reservoirs for

various microbial contaminants, including fungi, which pose significant risks to immunocompromised patients, healthcare workers, and visitors.

Fungal contaminants, such as *Aspergillus*, *Candida* and *Penicillium* species can enter the human body through inhalation and dermal contact, causing various reactions and symptoms in humans contributing to increased morbidity and mortality rates in healthcare settings<sup>1</sup>.

The presence of fungal contaminants on working surfaces in hospitals can be attributed to multiple factors, including poor ventilation, inadequate cleaning practices, and high foot traffic<sup>2,3</sup>. These fungi can be disseminated through air, water and direct contact leading to outbreaks that are challenging to control. This challenge may relate to the environment niches that they occupy; the relative ease of dispersal; the ability of fungi to grow on almost any substrate<sup>4</sup>, with the identification of fungi remaining a relatively specialist area when compared to the characterisation of bacteria

Fungi are ubiquitous in distribution and are a serious threat to public health<sup>5</sup>. Fungi are able to grow on almost all natural and synthetic materials, especially if they are hygroscopic or wet. Inorganic materials get frequently colonized as they absorb dust and serve as good growth substrates for *Aspergillus*<sup>6</sup>. Common contributors to biological pollutants are leaks in plumbing, roofs or air conditioners, humidifiers, and bathrooms; and ice damming on building roofs allows water to seep through the roof sheathing<sup>7</sup>. Fiber glass insulation and ceiling tiles support the growth of a number of fungi, among them frequently isolated were *A. versicolor*, *Alternaria*, *Cladosporium*, and *Penicillium* species<sup>8</sup>

This study aims to assess the incidence of fungal contaminants on various working surface areas in a tertiary teaching hospital in Amalapuram. By identifying the types and prevalence of fungi present, it is possible to understand the environmental factors contributing to fungal contamination and implement necessary infection control measures. The findings of this study will be instrumental in developing targeted strategies to reduce the risk of fungal infections and improve overall hospital hygiene and patient safety.

## **Objectives :**

The principal goal of this study is to identify the presence of fungi on various working surface areas of a Tertiary care Teaching Hospital, Amalapuram.

## **Materials and Methods :**

The present study was conducted in the Department of Microbiology, Konaseema Institute of Medical Sciences & Research Foundation, Amalapuram, from June 2023 to June 2024.

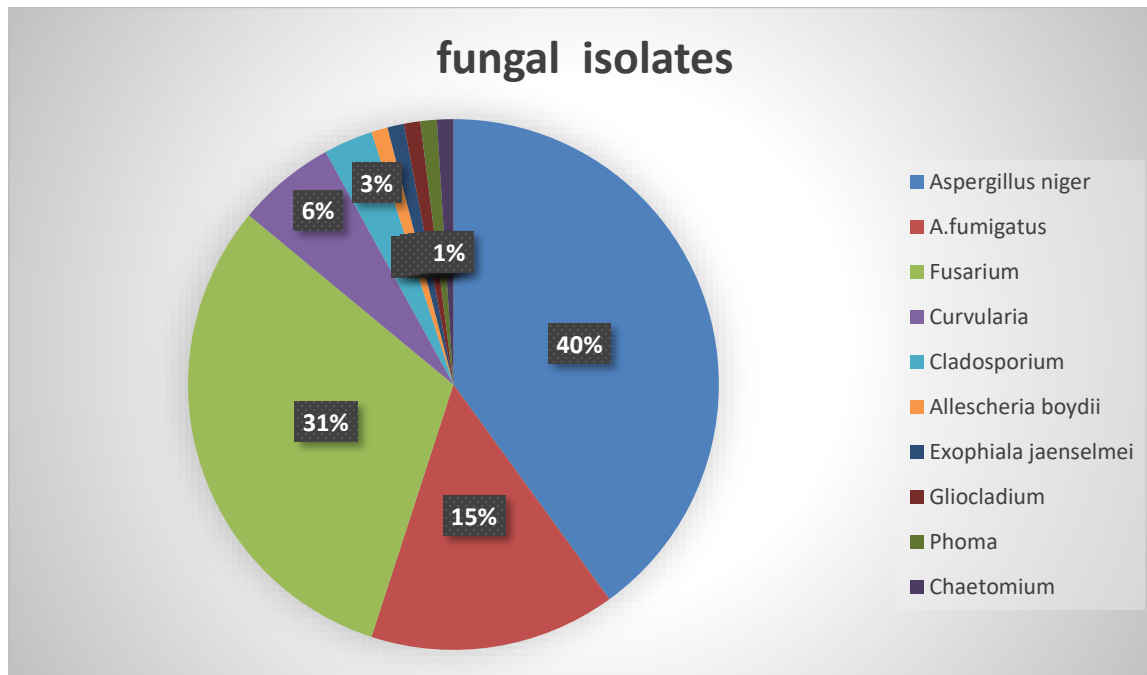
### **Sample collection method:**

- After obtaining Institutional Ethics Committee approval, samples were collected from various surface areas including bed rails, door handles, medical equipment, light switches, and nursing stations, ceilings, AC airvents, walls, IV stands etc...
- Surface samples were collected using sterile swabs moistened with sterile saline and were placed in sterile tubes and sent to microbiology lab as soon as possible.
- The swabs were inoculated onto Sabouraud's dextrose agar (SDA) with chloramphenicol to suppress any bacterial growth and incubated at 25<sup>0</sup> C in a BOD incubator temperature and checked daily for fungal growth for 7 days.
- Identification was made according to their macroscopic and microscopic morphological characteristics of the growth by standard mycological methods by Lacto phenol cotton blue (LCB) mount and other conventional tests.<sup>9,10</sup>

## RESULTS :

A total of 100 swabs were collected from different areas of the hospital- different wards, OTs, all ICUs. Out of 100 swabs, 64 swabs showed growth on SDA slants. Out of 64 isolates, 23 isolates were *Aspergillus niger* (40%), 10 were *A.fumigatus* (15%), 20 were *Fusarium* (31%), 4 were *Curvularia* (6%), 2 were *Cladosporium* (3%), *Allescheria boydii* -1(1%), *Exophiala jaenselmei*-1(1%), *Gliocladium* -1(1%), *Phoma* -1(1%), *Chaetomium* -1 (%)

Isolated Fungi	Total number	Percentage (%)
<i>Aspergillus niger</i>	23	40%
<i>A.fumigatus</i>	10	15%
<i>Fusarium</i>	20	31%
<i>Curvularia</i>	4	6%
<i>Cladosporium</i>	2	3%
<i>Allescheria boydii</i>	1	1%
<i>Exophiala jaenselmei</i>	1	1%
<i>Gliocladium</i>	1	1%
<i>Phoma</i>	1	1%
<i>Chaetomium</i>	1	1%



## DISCUSSION :

Moisture, nutrients and temperature are the most important factors that influence the growth of fungi on building materials (Rajasekar and Balasubramanian, 2011).<sup>11</sup>

Nutrients in house dust and water favor fungal growth on building materials. Fiberglass, galvanized steel accumulated with dust or lubricant oil residues, allows the growth of fungi (Kennedy et al., 2004, , Yau and Ng, 2011).<sup>12,13</sup> The temperature in buildings of about 20–25 °C, promotes the growth of mesophilic fungi. However, the temperature below optimum level slows down the growth of fungi. pH range of 5–6.5 in building materials allows the best growth of most of the fungi (Hoang et al., 2010).<sup>14</sup> Sufficient light and oxygen are also critical for the growth of fungi in indoor environments (Zadrzil et al., 1991, Airaksinen et al., 2004, Voisey, 2010).<sup>15,16,17</sup>

External factors (air humidity, seasonality, air and people flow, use of particulate filters, and health professionals' hand hygiene) contribute to indoor air contamination with fungi.

Improving communication among health professionals is a great concern because this can prevent major health complications. High infection rates may be attributed to working conditions, to the physical structure of the ward, to the nurse/bed ratio, to the resident microbiota in the care assistance, and to the presence of multiresistant microbial strains, among others (Jenyffie A et al., 2021).<sup>18</sup>

In the present study the most common isolates are of *Aspergillus* species (55%) which is correlating with other studies i.e Xunliang Tong et al., 2017 and in his study he worked at different critical care areas of hospital and concluded that 17 -61% of the fungi are of *Aspergillus* species.<sup>19</sup> In another similar study Dharendra Kumar et al., 2019 the most isolated fungi from

hospital environment and equipment are *Aspergillus* species (46.4%).<sup>10</sup> Similar study was also done by Parisa Badiie et al., 2023 and they stated that *Aspergillus* species (19%) are most commonly isolated fungal contaminants in hospital environment.<sup>20</sup> Jenyffie A et al., 2021 in a review of paper fungi in indoor area of critical area of hospital showed most of the fungal isolates in their study was *Aspergillus* species.<sup>18</sup> *Aspergillus* species cause aspergillosis infection usually affect the respiratory system and most serious form of aspergillosis is invasive aspergillosis where the infection spreads to blood vessels and beyond.<sup>21</sup>

The next common isolate in our study is *Fusarium* (31%) while in other studies Dhirendra Kumar et al., 2019 and Parisa Badiie et al., 2023 *Fusarium* isolates are 7%.<sup>10,20</sup> In a review paper by Jenyffie A et al., 2021 it was clear that *Fusarium* is also one of the most common fungal isolate in hospital environment next to *Aspergillus* species.<sup>18</sup> *Fusarium* generally cause keratitis and onychomycosis. It may also cause allergic diseases (sinusitis) and mycotoxicosis in humans.<sup>22,23</sup>

In the present study *Curvularia* spp (6%) has been isolated next to *Fusarium* and it has been also isolated in other studies like Frequency of airborne fungus in critical areas at hospital unit by Venceslau et al., (2012) and other study Fungal biodiversity of air in Hospitals Pantoja et al., (2012).<sup>24,25</sup> *Curvularia* are human pathogens which cause mild skin and nail infections to severe invasive disease.<sup>26</sup>

There are *Cladosporium* (3%) isolates in our study and these isolates are also seen in 0.5% in Parisa Badiie et al., 2023 study and in Hoseinzadeh et al. (2013) Evaluation of bioaerosols in five educational hospitals wards *Cladosporium* is a common isolate.<sup>20,27</sup> It is also seen in Calumby et al. (2019) Isolation and identification of anemophilic fungal microbiota in ICU and Souza et al (2019) Airborne fungi in neonatal intensive care unit of a public hospital as a most common fungal isolate.<sup>28,29</sup>

Aboul-Nasr et al. (2014) Indoor surveillance of airborne fungi contaminating Intensive care units and operation rooms and El-Sharkawy et al (2014) Indoor air quality levels in a University Hospital also stated that *Cladosporium* one of the fungal isolates in their study.<sup>30,31</sup> *Cladosporium* has been reported to cause several different types of opportunistic infections, including subcutaneous and deep infections, in humans. *Cladosporium* spores, also potentially lead to the development of respiratory allergy problems such as asthma and rarely cause pulmonary infection.<sup>32</sup>

In the present study other rare isolates are *Allescheria boydii* (1%), *Exophiala jaenselmei* (1%), *Gliocladium*(1%), *Phoma*(1%) and *Chaetomium*(1%). In Dhirendra Kumar et al., 2019 study also *Exophiala jaenselmei* (1.8%) and *Phoma*(1.8%) were isolated.<sup>10</sup>

*Allescheria boydii* & *Exophiala jaenselmei* can cause mycetoma.<sup>33</sup> *Gliocladium* rarely cause ocular infections.<sup>34</sup> *Phoma* causes rare opportunistic infections like subcutaneous mycoses and onychomycosis.<sup>35</sup> *Chaetomium* can cause sinusitis, onychomycosis and empyema.<sup>36</sup>

## CONCLUSION:

Most of the patients in the hospitals are immune compromised. These fungal isolates in the hospital environment may become an additional economic burden to their present situation as most of them are multi drug resistant. So, measures to control hospital infection in the hospital environment mainly ICUs is very much needed. So it is necessary to bring awareness that there is a chance of acquiring fungal infections as they commonly form spores and spread infections. Hence promoting good air ventilation and periodical cleaning of air conditioners to control excess moisture in the walls and hospital environment to decrease the fungal saprophytes according to standard protocols is necessary. Also alerting about the situation of hospital environmental fungal infections to the health professionals and hospital management is a crucial step in infection control.

## REFERENCES :

1. Ramos CA, Viegas C, Verde SC, Wolterbeek HT, Almeida SM. Characterizing the fungal and bacterial microflora and concentrations in fitness centres. *Indoor Built Environ.* 2015;**25**(6):872–882.
2. Bauer TM, Ofner E, Just HM, Just H, Daschner FD. An epidemiological study assessing the relative importance of airborne and direct contact transmission of microorganisms in a medical intensive care unit. *J Hosp Infect.* 1990;**15**(4):301-9.
3. Kramer A, Schwebke I, Kampf G. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. *BMC Infect Dis.* 2006;**6**:13
4. Hawksworth DL, Lücking R (July 2017). Fungal Diversity Revisited: 2.2 to 3.8 Million Species. *The Fungal Kingdom - Microbiology Spectrum.* 5. pp. 79–95
5. Z.U. Khan, M.A.Y. Khan, R. Chandy, P.N. Sharma, *Aspergilli* and other moulds in the air of Kuwait, *Mycopathologia*, 146 (1) (1999), pp. 25-32
6. J.M. Samet, J.D. Spengler -Indoor environments and health: Moving into the 21st century *American Journal of Public Health*, 93 (9) (2003), pp. 1489-1493.
7. M.J. Cunningham, C. Roos, L. Gu, G. Spolek, Predicting psychrometric conditions in biocontaminant microenvironments with a microclimate heat and moisture transfer model description and field comparison, *Indoor Air*, 14 (2004), pp. 235-242
8. I.P. Erkara, A. Asan, V. Yilmaz, S. Pehlivan, S.S. Okten, Airborne *Alternaria* and *Cladosporium* species and relationship with meteorological conditions in Eskisehir City, Turkey, *Environmental Monitoring and Assessment*, 144 (1–3) (2008), pp. 31-41
9. Garcia-Cruz CP, Najera Aguilar MJ, Arroyo-Helguera OE. Fungal and Bacterial Contamination on Indoor Surfaces of a Hospital in Mexico. *Jundishapur J Microbiol.* 2012;**5**(3):460-4. DOI: 10.5812/jjm.262
10. Dharendra Kumar Nahid\_Anjum Sayan B ISOLATION OF FUNGUS FROM HOSPITAL ENVIRONMENT AND EQUIPMENTS IN A TERTIARY CARE HOSPITAL IN EASTERN INDIA Volume-8 | Issue-2 | February-2019 | PRINT ISSN No 2277 - 8179
11. Rajasekar, R. Balasubramanian, Assessment of airborne bacteria and fungi in food courts, *Building and Environment*, 46 (10) (2011), pp. 2081-2087
12. S.M. Kennedy, R. Copes, K.H. Bartlett, M. Brauer, Point-of-sale glass bottle recycling: indoor airborne exposures and symptoms among employees, *Occupational and Environmental Medicine*, 61 (2004), pp. 628-635

13. Y.H. Yau, D. Chandrasegaran, A. Badarudin, The ventilation of multiple-bed hospital wards in the tropics: a review, *Building and Environment*, 46 (5) (2011), pp. 1125-1132
14. C.P. Hoang, K.A. Kinney, R.L. Corsi, P.J. Szaniszló, Resistance of green building materials to fungal growth, *International Biodeterioration Biodegradation*, 64 (2) (2010), pp. 104-113
15. F. Zadrazil, G.C. Galletti, R. Piccaglia, G. Chiavari, O. Francioso, Influence of oxygen and carbon dioxide on cell wall degradation by white-rot fungi *Animal Feed Science and Technology*, 32 (1–3) (1991), pp. 137-142
16. M. Airaksinen, P. Pasanen, J. Kurnitski, O. Seppanen, Microbial contamination of indoor air due to leakages from crawl space. A field study, *Indoor air*, 4 (2004), pp. 55-64
17. C.R. Voisey, Intercalary growth in hyphae of filamentous fungi *Fungal Biology Reviews*, 24 (3–4) (2010), pp. 123-131
18. Jenyffie A. Belizario . Leonardo G. Lopes . Regina H. Pires Fungi in the indoor air of critical hospital areas: a review *Aerobiologia* (2021) 37:379–394 [https://doi.org/10.1007/s10453-021-09706-7\(0123456789\(\).,-volV\)\( 01234567](https://doi.org/10.1007/s10453-021-09706-7(0123456789().,-volV)( 01234567)
19. XunliangTong1,\* , HongtaoXu2,\* , LihuiZou3, MengCai4, XuefengXu5, Zuotao Zhao6, FeiXiao3 & Yanming Li4,7 High diversity of airborne fungi in the hospital environment as revealed by meta-sequencingbased microbiome analysis *Scientific Reports* | 7:39606 | DOI: 10.1038/srep39606
20. Parisa Badiee,1 Abdolkarim Ghadimi-Moghadam,2 Habibeh Bayatmanesh,2 Jafar Soltani,3 Ali Reza Salimi-Khorashad,4 Fatemeh Ghasemi,1 Maneli Amin Shahidi,1 Hadis Jafarian1 Environmental surveillance of fungi and susceptibility to antifungal agents in tertiary care hospitals <https://journals.asm.org/journal/spectrum> on 17 September 2024 by 2401:4900:6760:d1e0:6160:995b:cc88:f899
21. Lajonchere JP, Feuilhade de Chauvin M. Contamination by aspergillosis: evaluation of preventive measures and monitoring of the environment [in French]. *PatholBiol (Paris)* 1994; 42:718-729.
22. Guarro J. Fusariosis, a complex infection caused by a high diversity of fungal species refractory to treatment. *Eur J Clin Microbiol Infect Dis*. 2013;32(12):1491–1500.
23. Dabas Y, Bakhshi S, Xess I. Fatal cases of bloodstream infection by *Fusarium solani* and review of published literature. *Mycopathologia*. 2016;181(3-4):291–296.
24. Venceslau, E. M., Martins, R. P. P., & Oliveira, I. D. (2012). Frequency of airborne fungus in critical areas at hospital unit of Aracaju, Sergipe, Brazil. *Revista Brasileira Analises Clinicas*, 44(1), 26–3
25. Pantoja, L. D. M., Couto, M. S., Junior, N. P. L., de Sousa, B. L., Moura, C. I., & Paixão, G. C. (2012). Fungal biodiversity of air in hospitals in the city of Fortaleza, Ceará, Brazil *Revista Brasileira de Promoc,ão da Sau´de*, 25(2), 192–196
26. Madrid H, da Cunha KC, Gené J, Dijksterhuis J, Cano J, Sutton DA, et al. Novel *Curvularia* species from clinical specimens. *Persoonia*. 2014;33:48–60
27. Hoseinzadeh, E., Samarghandie, M. R., Ghiasian, S. A., Alikhani, M. Y., & Roshanaie, G. (2013). Evaluation of bioaerosols in five educational hospitals wards air in Hamedan, during 2011–2012. *Jundishapur Journal of Microbiology*, 6(6), e10704

28. Calumby, R. J. N., Silva, J. A., Moreira, R. T. F., Araujo, M. A. S., & Almeida, L. M. (2019). Isolation and identification of anemophilic fungal microbiota in an intensive care unit. *Brazilian Journal of Development*, 5(10), 19708–19722\
29. Souza, A. K. P., Nascimento, J. P. M., Araujo, M. A. S., Pedrosa, K. P. S., Tenorio, B. M., Pires, L. L. S., et al. (2019). Airborne fungi in neonatal intensive care unit of a public hospital in Brazil. *International Journal of Current Microbiology and Applied Sciences*, 8(12), 1210–1219
30. Aboul-Nasr, M. B., Abdel-Naser, A. Z., & Amer, E. M. (2014). Indoor surveillance of airborne fungi contaminating intensive care units and operation rooms in Assiut University hospitals. *Egypt Journal of Health Science*, 2, 20–27
31. El-Sharkawy, M. F., & Noweir, M. E. (2014). Indoor air quality levels in a university hospital in the Eastern Province of Saudi Arabia. *Journal of Family & Community Medicine*, 21(1), 39–47
32. Crous PW, Braun U, Schubert K, Groenewald JZ. 2007. Delimiting *Cladosporium* from morphologically similar genera. *Stud Mycol* 58:33–56. doi: 10.3114/sim.2007.58.02
33. Kwon-Chung KJ, Bennett JE. *Medical Mycology*. Philadelphia: Lea & Febiger; 1992. Mycetoma; pp. 560–93
34. Venkatesh R, Gurav P, Agarwal M, Sapra N, Dave PA. Ocular infection with *Gliocladium* species-report of a case. *J Ophthalmic Inflamm Infect*. 2017 Dec;7(1):9. doi: 10.1186/s12348-017-0128-1. Epub 2017 Mar 14. PMID: 28293854; PMCID: PMC5350082
35. Bennett A, Ponder MM, Garcia-Diaz J. Phoma Infections: Classification, Potential Food Sources, and Its Clinical Impact. *Microorganisms*. 2018 Jun 23;6(3):58. doi: 10.3390/microorganisms6030058. PMID: 29937481; PMCID: PMC6165263
36. Kim DM, Lee MH, Suh MK, Ha GY, Kim H, Choi JS. Onychomycosis Caused by *Chaetomium globosum*. *Ann Dermatol*. 2013 May;25(2):232-6. doi: 10.5021/ad.2013.25.2.232. Epub 2013 May 10. PMID: 23717019; PMCID: PMC3662921